Pacific Community
Communauté du Pacifique

BEST PRACTICES

for the collection, transport, holding and export of fish and corals in the aquarium trade

Colette Wabnitz
Tony Nahacky

FAME
Fisheries, Aquaculture and Marine Ecosystems Division

BEST PRACTICES

for the collection, transport, holding and export of fish and corals in the aquarium trade

Colette Wabnitz
Fisheries Scientist (Aquarium trade), SPC
*current affiliation: Institute for the Oceans and Fisheries, University of British Columbia, Vancouver, Canada

Tony Nahacky
Independent consultant

Pacific Community
Communauté du Pacifique

Noumea, New Caledonia, 2019

Original text: English

Pacific Community Cataloguing-in-publication data

Wabnitz, Colette

Best practices for the collection, transport, holding and export of corals and fish in the aquarium trade / Colette Wabnitz and Tony Nahacky

1. Corals.
2. Fishes.
3. Aquarium fish collecting.
4. Aquarium fishes.

I. Wabnitz, Colette II. Nahacky, Tony III. Title IV. Pacific Community

639.34 AACR2

ISBN: 978-982-00-1193-9

This document may be cited as:

Wabnitz C. and Nahacky T. 2019. Best practices for the collection, transport, holding and export of fish and corals in the aquarium trade. Noumea, New Caledonia: Pacific Community. 18 p.

Technical illustrations by Jipé Le-Bars, SPC
Layout and graphics by Boris Colas, SPC.
Cover picture: *Pomacanthus imperator* by Brian Gratwicke, Flickr

Prepared for publication by the Pacific Community, Noumea, New Caledonia – www.spc.int

INTRODUCTION

These guidelines were compiled and based on what is currently considered as "best practice" in the collection, transport, holding and export of marine aquarium organisms that are destined for the hobby trade, rather than, for example, public aquaria. However, and perhaps most importantly, they also describe procedures that will hopefully contribute to the long-term sustainability of local resources, which, in turn, is essential for sustaining the local trade and economy.

While these practices are based on the best available information at the time of writing, are informed through experience, and fine-tuned to best fit a range of local conditions and settings in the Pacific Islands region, they cannot possibly be, and are not meant to be, fully comprehensive. There are many means to ensure that fish and corals are collected, held and shipped in perfect conditions and arrive alive and in good health. Here, we have attempted to compile – as succinctly as possible – suggestions on how to achieve this without being prescriptive.

In addition to the recommendations outlined in this publication, businesses should always comply with relevant regulations and adhere to local labour laws.

Implementation of these practices should be accompanied by a management plan that is specifically designed for regulating marine aquarium fishery practices/activities, and include regulations that limit the number of licences and permits issued for operators/collectors within given areas.

SAFETY for diving collectors[1]

- Collectors using any type of underwater breathing apparatus will need to be certified for the equipment they are using (e.g. scuba) and provide a copy of the certificate to the exporter and, if applicable, the relevant (local) authority.
- Divers must have and use adequate and safe diving equipment – and maintain these properly.
- **As recommended by the American Association of Underwater Science and the Divers Alert Network, start a dive, or a series of dives, at the maximum planned depth, followed by subsequent dives of shallower exposures (i.e. do your deepest dive first and plan for other dives in shallower water).**
- **Divers should ensure they have the appropriate rest time/surface interval between dives – spend as much time on the boat before your next dive as recommended by dive tables and/or your dive computer.**
- All gear and equipment, including boats and compressors, should comply with generally recognised and accepted health and safety criteria (e.g. seaworthiness of boats, regular servicing and testing of scuba tanks and compressor, etc.).

[1] These are general safety guidelines and are not meant to be fully comprehensive. For safety procedures, please consult the Divers Alert Network (www.dan.com) or the American Academy of Underwater Sciences (http://www.aaus.org/mc/page.do) on a regular basis.

FISH

Verify order (importer > exporter > collector); and stick to collecting species and numbers on order list. Instructions must include how many individuals/pieces and of which species and size. Inexperienced collectors should learn to identify the fish they are to collect from books, the internet, the knowledge of experienced fishers, etc.

Based on the order, the buyer (importer and exporter) needs to be specific about which sizes and/or colours that collectors should not harvest (i.e. does not wish to receive).

Keep records of how many fish were caught, their species, and where and by whom they were caught.

Provide appropriate records of all exports to the relevant local authorities, as specified by the local authorities (e.g. typically, at a minimum, the records should include quantities exported, by species, shipment date and shipment destination).

Do not use poison of any kind (synthetic or plant derived chemicals) to catch fish and/or other animals.

Ensure collection of fish and other animals does not damage the reef habitat or create waste. If you have moved rocks, return them to their original position.

When anchoring at a collection site avoid damage to the reef habitat.

Rotate collection areas.

Utilise mesh of the right/appropriate size for targeted species to avoid gilling of and/or damaging animals (typically mesh size should be 20 mm or smaller).

Maintain all netting equipment in good condition, and patch holes as soon as they occur to avoid gilling.

Always attend to your nets – never leave nets unattended, underwater.

Minimise touching of the fish at all times – if necessary, ensure you have clean hands and then handle the fish by gently grabbing it around the gill plate (not the softer main body).

Fish should be collected with enough time before the packing date to allow the animals to "purge" and clean out their stomachs. Fish that have, for some reason, been kept in cups at sea or at a holding facility without being fed for longer than the fasting times indicated in Table 1 should be returned to the sea.

Table 1

Fish size	No. of days	Max. no. days fasting
Small (< 2 in or < 5 cm)	2–3 days	5 days
Medium (2–4 in or 5–10 cm)	3–4 days	6 days
Large (4–6 in or 10–15 cm)	4–6 days	7 days

When collecting fish underwater, do not exceed the number of fish in a 7 L container (Fig. 1) than specified in Table 2.

Figure 1

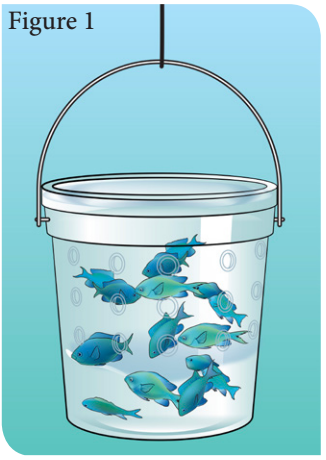
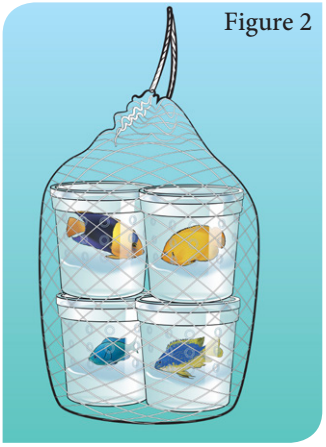


Table 2

Fish size	Max. no. of fish
Small (< 2 in or < 5 cm)	20–25 fish (3–4 fish per L)
Medium (2–4 in or 5–10 cm)	12–15 fish (2 fish per L)
Large (4–6 in or 10–15 cm)	8–10 fish (1 fish per L)

Do not collect fish bigger than 6 in or 15 cm.

Figure 2



For angelfish and other species that fight with other fish – e.g. filefish, triggerfish, tomato/cinnamon clowns, hawkfish and all *Centropyge* species – place the fish inside a cup, and then place all the cups holding fish in a soft net and hang it off at the appropriate depth to allow for the fish to decompress (Fig. 2).

Ensure individual fish containers/cups have sufficient holes, and that the inside of the cup is smooth. Do not put any holes in the bottom of the cups or the bottom third of the sides of the cup.

Decompression techniques

Use appropriate fish decompression techniques.

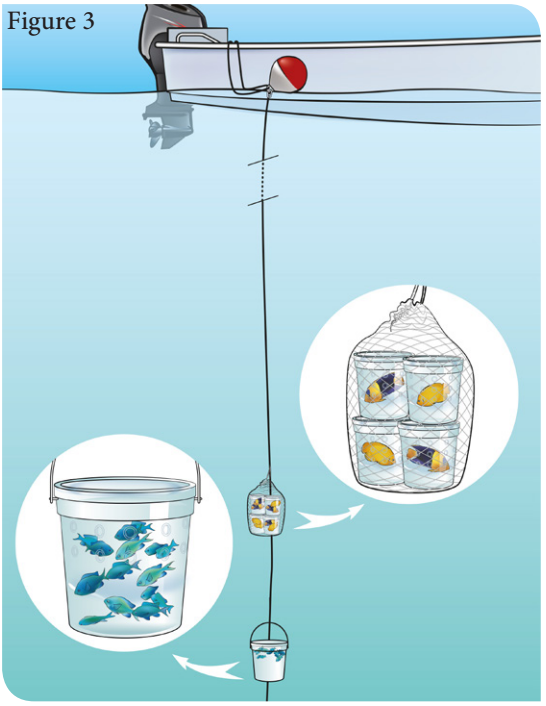
Before starting a dive, attach a rope with a weight suspended within 5 m from the sea floor and a float at the top of the line to your boat.

A combination of the techniques detailed below can be used. Always monitor the behaviour of the fish, and only needle the fish that are struggling to swim and/or hanging at a steep angle (vertically) in the water.

Fish decompression by gradual depth change

At the end of the dive, ascend slowly and watch the fish. The fish should be swimming easily, at a slight angle; they should not be struggling. As soon as one of the fish in the bucket swims at a 45° angle, hang the bucket off the rope (Fig. 3).

Figure 3



Check on the fish every 30 to 60 minutes or more frequently. If the fish are swimming easily (Fig. 4), raise the bucket until one fish starts swimming at a 45° angle, and hang the bucket off the rope again.

Proceed in this manner until you reach the surface.

If a fish is having a lot of trouble swimming and is hanging vertically in the bucket, it is decompressing too close to the surface (i.e. too shallow) and the bucket should be lowered.

The use of a clip or a carabiner for hanging the bucket containing the fish off the rope (to decompress the fish) is the preferred method.

Fish decompression by needling

Keep needles in alcohol to minimise infection to the fish.

Make sure that the needle is clear (by blowing water through the needle – be careful when handling the needle; or if possible use a syringe to squirt water through the needle).

Make sure the needle is not bent or kinked. If it is, dispose of it appropriately (i.e. somewhere safe). Replace needles regularly.

Insert the needle into the fish, under the scales, at a slight angle and only push in very slightly and gently (i.e. do not push on the needle perpendicularly to the fish's body; do not push the needle through the fish).

Do not squeeze the fish while needling as too much air will come out.

Fish can be needled through the anus or the side; choose whichever method produces the healthiest fish and minimises infection risks. For example, angelfish, butterflyfish and surgeonfish (tangs) are typically needled through the side, while anthias and wrasses are needled through the anus.

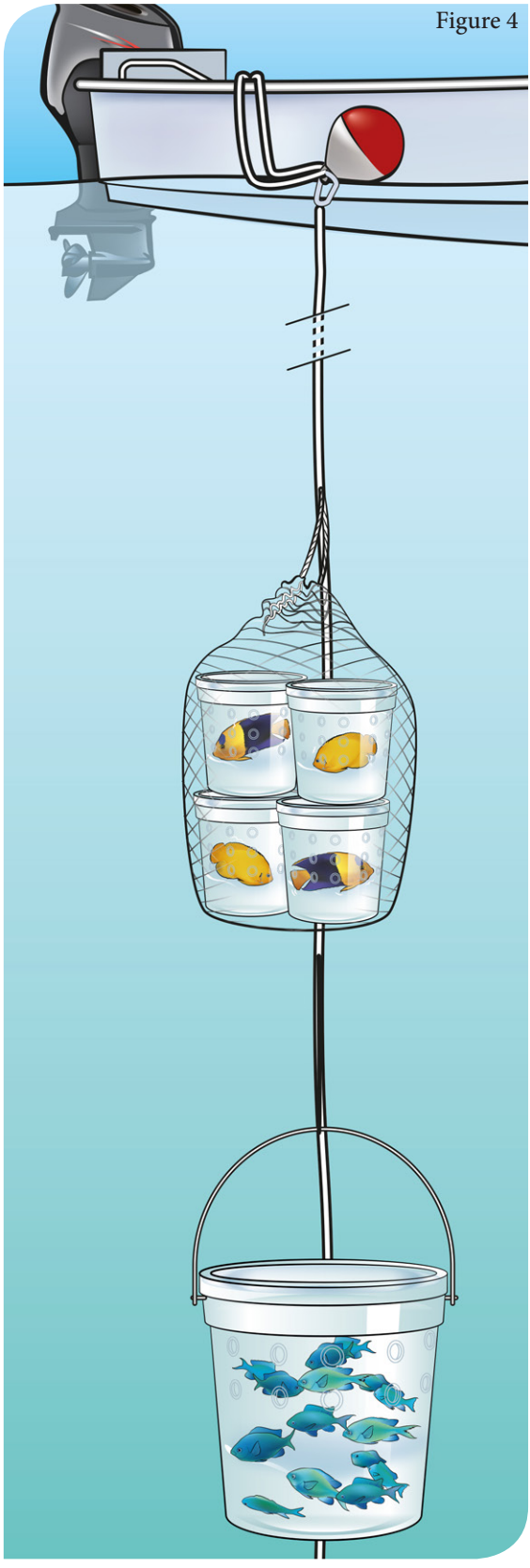
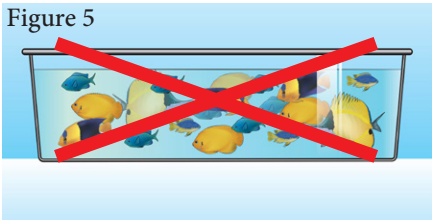


Figure 4

The container/bin for fish on the boat (e.g. large bucket, or Rubbermaid tub) needs to be big enough to not crowd the fish and to maintain sufficient water so the fish have room to swim (Fig. 5). However, very tall containers such as large garbage tubs should be avoided[2].

Figure 5



On the boat, if you do not have a flow system that provides fish with clean water automatically and regularly, do not exceed the number of fish in a 35 L container (e.g. Rubbermaid), as specified in Table 3.

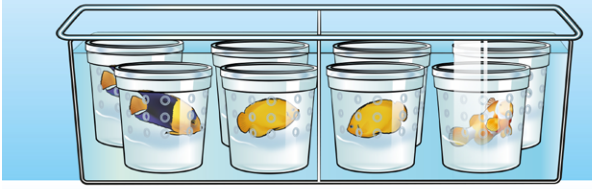
Table 3

Fish size	Max. no. of fish
Small (< 2 in or 5 cm)	30–35 fish (0.5–1 L per fish)
Medium (2–4 in or 5–10 cm)	12–17 fish (1–2 L per fish)
Large (4–6 in or 10–15 cm)	7–11 fish (2–4 L per fish)

Water depth should not exceed 24 in or 60 cm.

Cups inside a fish-holding container must always be full of water and submerged (Fig. 6).

Figure 6



Species that are known to fight with other fish (e.g. filefish, triggerfish, tomato/cinnamon clowns, hawkfish and all *Centropyge* species) should be separated through compartments or kept in the separate cups into which they were placed underwater.

Rough fish (such as triggerfish and surgeonfish) should be kept separately from soft scaled fish.

[2] The reason being that the fish will stay at the bottom, but oxygen cannot get to the bottom, especially if tubs are filled with water > 60 cm depth, so the fish will run out of oxygen.

All boxfish, pufferfish, soapfish and other toxic fish (e.g. *Diploprion bifasciatum*, *Grammistes sexlineatus*) should not be kept in the same container/bin as other fish – each fish should be kept separately.

Containers should be covered to protect fish from the sun (and always placed in the shade when possible) and to prevent any material (food, ashes, fuel) from getting into the water.

Water changes need to be done every 30 to 45 minutes when fish are kept at the recommended density in Table 3. This is to ensure adequate levels of oxygen and a constant temperature are maintained in the holding bucket.

When performing a water change, always leave enough water in the container for fish to swim easily (i.e. do not remove water so the fish start swimming on their side).

Inspect all containers before leaving the collection site. Injured or deformed animals should be returned to the sea at the collection site prior to returning to harbour.

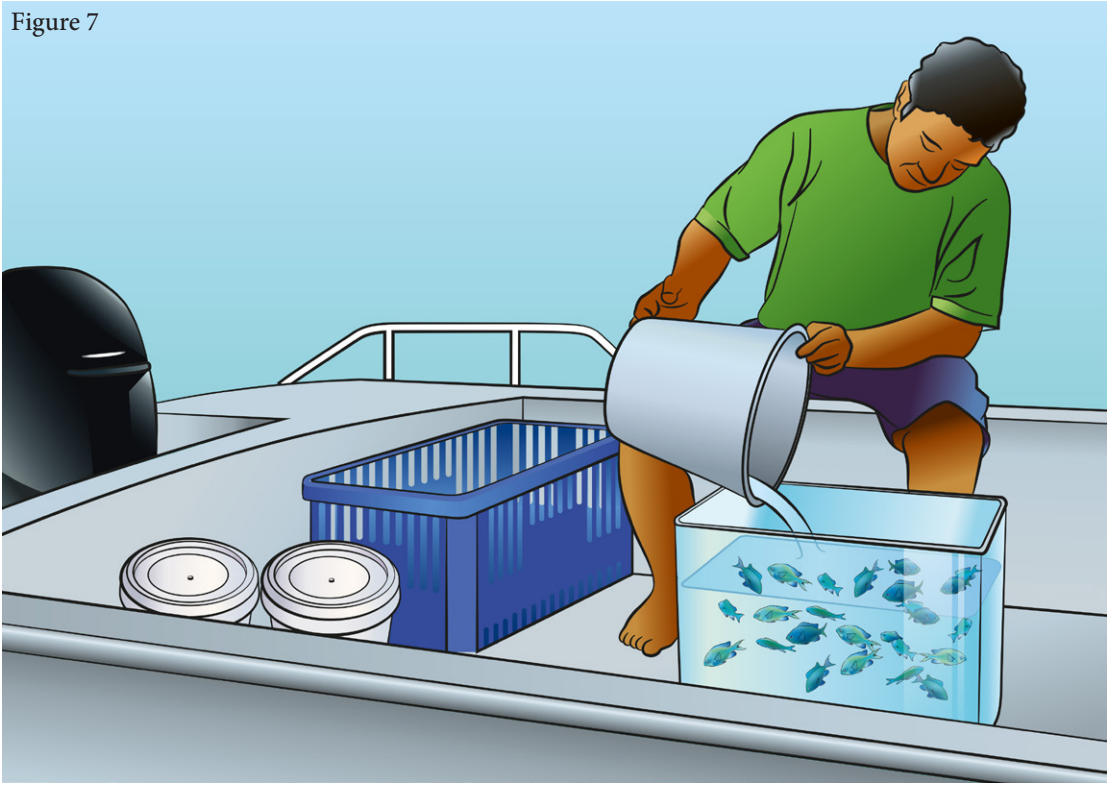
Do not put containers/bins with fish in the front of the boat as this is the location on a boat that is most likely to cause sloshing, which damages the fish.

When the boat is moving, minimise sloshing. If water splashes out of the bin while the boat is moving, stop and fill the bin back up.

Minimise the amount of time the fish will spend in the bin on the boat; get the fish back to the facility as soon as possible.

Before coming into the harbour, perform a water change and make sure this is done in an area with clean water (Fig. 7).

Figure 7

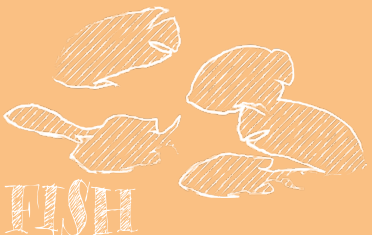


If the time from the last water change to the delivery at the facility is less than 30 minutes, remove some water to make transport easier. After partial water removal there should be, at minimum, enough water in the bin for all fish to be covered when in a vertical/upright position. If bins contain cups, after water removal all cups should still be full of water.

If the time from the last water change to the delivery at the facility is greater than 30 minutes, do not remove water for transport and ensure sufficient aeration or fresh seawater is provided.

Transfer one fish at a time (in its cup if applicable). Always have the bottom third of the cup full of water.

For fish that are not in a cup, transfer fish with a soft hand net. Minimise the amount of time the fish is out of water when using the net.



Fish should be kept in adequate land-based facilities.

The following list should be printed, regularly checked and steps should be taken to ensure all statements are true:

- Facility is regularly cleaned (including all holding containers) and good hygiene is undertaken to minimise the spread of disease and the risk of infestation of insects, rats and mice.
- The filtration system is maintained properly.
- Water purity is checked by chemical analysis using commercially available test kits.
- The facility has sterilisation measures to control bacteria/disease (e.g. UV lights, making sure bulbs are replaced regularly).
- Water temperature is measured regularly and always falls within the prescribed limits to maintain optimal health of the organisms.
- Adequate water flow is provided throughout the facility – a minimum four times turnover an hour within the container for fish.
- Animals are inspected at least daily.
- Fish cubicles or cups are clean.
- There is an indication of collection day for the fish in the facility.

A fish should always be allowed enough room to swim in circles/turn around in its holding container.

Species that are kept together (e.g. schooling fish) should not be overcrowded.

Species that are known to fight with other fish (e.g. angelfish, filefish, triggerfish, tomato/cinnamon clowns, hawkfish and all *Centropyge* species) should be kept in separate containers (i.e. one fish per cup/cubicle).

All boxfish, pufferfish, soapfish (e.g. *Diploporium bifasciatum*, *Gramistes sexlineatus*) should be kept separately from all other fish as they are poisonous and can kill other fish.

Allow for plenty of water exchange for each container (i.e. if a fish is held in a cup, the cup should have enough holes to allow for water exchange).

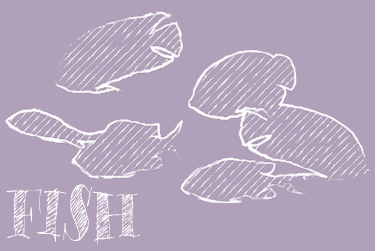
When making holes in cups make sure the inside is smooth to minimise damage to the fish. Do not place holes in bottom of cup or lower third of the cup's sides.

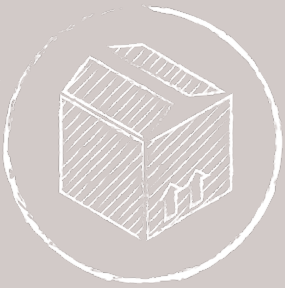
Temperature control

Maintain fish at the optimal temperature, which is between 74°F and 80°F (23°C and 26.6°C) for recirculating systems.

For flow through (open) systems, place intake pipe deep enough below the surface to maintain water between 74°F and 80°F (23°C and 26.6°C), aiming for the middle of the range.

Keep a log of mortalities. Dispose of dead animals in a safe and hygienic manner.





Make sure all packing materials and shipping documents are accessible and filled out as much as possible before the pack starts.

Make sure that all packing materials and the packing area are clean.

Make sure hands are washed prior to packing, paying particular attention to remove/scrub away any oil or dirt and make sure to rinse hands well so no soap is left behind.

Do not pack fish before you have to, in order to ensure optimal health (i.e. do not pack animals earlier than needed).

Do not overcrowd fish during the packing process.

Do not pack fish that have just been collected – the fish's waste inside the bag will affect its health during transport.

Always screen organisms prior to export, making sure they are in good physical condition (i.e. fins, eyes, body are looking healthy) and behaving normally.

Any animal that looks weak or is infected with parasites and bacteria should not be dispatched (e.g. rotten/melting fins, cloudy fins, spots on the fish, missing scales, cloudy eyes, pop eye, high respiration rate, embedded parasites, swollen stomach, red anus, red streaks on body, struggling to stay down or struggling to stay up).

Keep physical contact with fish to an absolute minimum.

Packing water should be uncontaminated – i.e. collected from a clean ocean site. Containers not in use should be covered at all times.

At a minimum, always double bag. For fish species with sharp spines and teeth, more bags or thicker bags are recommended.

Have clear liner covering the bottom portion of the bag.

Use the following guidelines (Table 4) for optimal packing of fish – keeping in mind to:

- always only pack one live fish/animal per bag;
- make sure that the fish can turn around in the bag;
- choose a bag that is at least twice the body length of fish in width; and
- fill water in the bag so the top of the dorsal fin is underwater (i.e. enough water to cover the fish and about an inch (2.5 cm) more).

Table 4

Fish size	Bag size
Small (< 2 in or 5 cm)	4–6 in or 10–15 cm
Medium (2–4 in or 5–10 cm)	7–9 in or 17–22 cm
Large (4–6 in or 10–15 cm)	7–11 in or 17–27 cm

For packing of aquarium organisms, carry out the following:

- Select a double bag and fill it with packing water to a level appropriate for the size of the organism to be placed into the bag.
- Gently place the organism into the bag.
- Make sure the animal is fully submerged (i.e. for fish enough water to cover the fish and about an inch (2.5 cm) more).
- Take all the air out of the bag before adding oxygen.
- Add sufficient oxygen to the bag – make sure that the fish has sufficient water and oxygen, and remember that larger fish will need more of both.
- Twist the bag prior to sealing it (Fig. 8) – it is best practice to twist only the top portion of the bag (i.e. the plastic that is to be banded/clipped) and not the entire bag, but if you do then do so *slowly and gently*; if done quickly it may stress the animal inside or/and puncture the bag.
- Seal the bag tightly.
- Line a box with protective plastic liner to retain any potential leakage.
- Insulate the box (e.g. styrofoam box or if styrofoam is not available two boxes separated by crumpled, stuffed newspaper).

Figure 8

Do not pack fish in water outside of the holding range (between 74°F and 80°F (23°C and 26.6°C)).

Always maintain bags upright.

Fill empty spaces in boxes with air-filled plastic bags.

If you are putting two layers of bags in a box, ensure that adequate support is provided for that second layer. In other words, ensure that the weight of the second layer is not resting on the first layer of bags, by placing a structure to hold up the second layer.

Ensure your shipment meets all the requirements prior to export, including but not limited to:

- 48-hour notification to importers;
- detailed packing list (quantities by species in each box);
- summary invoice;
- export permits; and
- copy of airway bill – with flight number and estimated time of arrival (ETA).

Label all boxes appropriately – species, number of animals, LIVE FISH, UP arrow, temperature, shipper's address and consignee address (i.e. who the shipment is going to).

Always adhere to the International Air Transport Association's (IATA) and the specific airline's policies.

IATA specifies that fish should be packed for up to a 24-hour delay. If the departure flight is delayed by close to or more than 24 hours, all fish will need to be brought back to the facility and repacked for the new flight schedule once it has been confirmed.

If the plane is delayed by less than 24 hours and the fish have not been packed to accommodate up to the 24-hour delay as recommended by IATA, the fish will need to be brought back to the facility and repacked for the new flight schedule.

Always keep boxes in a cool, shaded place. Do not store and/or transport packed boxes under the sun.

Total shipping time between exporter (beginning of pack) and importer (last specimen out of its bag) should ideally be 24 hours (up to 48 hours).

Always make sure to bring originals and copies of permits and other shipment documents to the airport.

If a connecting flight is needed between origin and destination, make sure that a backup flight to the same origin or destination is available within 24 hours of the original connecting flight – meaning your animals can get to your importer or sent back to you within 24 hours. If this is not the case, no live animals should be shipped to that destination.

CORALS

Verify the order (importer > exporter > collector); and stick to collecting species and numbers on the order list. Instructions must include how many individuals/pieces and of which species. Inexperienced collectors should learn to identify corals they are to collect from books, the internet, the knowledge of experienced fishers, etc.

Based on the order, the buyer (importer and exporter) needs to be specific about which sizes and/or colours that collectors should not harvest (i.e. does not wish to receive).

Keep records of how many corals were caught, their species, and where and by whom they were caught.

Provide appropriate records of all exports to the relevant local authorities, as specified by the local authorities (e.g. typically, at a minimum, the records should include quantities exported, by species, shipment date and shipment destination).

Ensure collection of corals does not damage the reef habitat or create waste. If you have moved rocks, return them to their original position.

When anchoring at a collection site avoid damage to the reef habitat.

Rotate collection areas.

Harvest all attached corals using a small hammer and chisel.

For collection, use a container that minimises rolling of individual coral pieces.

Avoid stacking of the corals (i.e. putting corals on top of each other).

Before collecting any coral, first ask these questions:

- Is this coral on the order?
- Is this the right kind of coral?
- Is it the right size of coral?
- Is it the right colour of coral?
- Is it the right shape of coral?
- Can the coral be collected without breaking it?
- Is it free of imperfections/dead spots?

If the answer is YES to ALL of the above, then collect the coral.

If the answer is NO to ANY question above, do not collect the coral.

Do not harvest large coral colonies (> 5 in/13 cm).

Collected corals should be placed in one layer in a shallow Rubbermaid container filled with fresh seawater.

Each coral piece should be separated by the appropriate amount of space from the next piece – about the equivalent of two fingers.

If corals have to be stacked, form a maximum of two layers. The two layers will need to be separated by plastic or some other form of soft material.

Hard and soft corals should be maintained in separate containers, according to species, to ensure they are not in direct contact with each other.

Minimise exposure to air, direct sunlight, and temperature extremes.

Change water regularly – every 30 to 45 minutes.

If some species are producing a large amount of mucus or colouring the water, perform a water change sooner.

If the time from the last water change to the delivery at the facility is less than 30 minutes, remove some water to make transport easier. After partial water removal there should be enough water in the bin for all corals to be covered.

If the time from the last water change to the delivery at the facility is greater than 30 minutes, do not remove water for transport and ensure sufficient aeration or fresh seawater is provided.

Corals should be kept in adequate land-based facilities.

The following list should be printed, regularly checked and steps taken to ensure all statements are true:

- Facility is regularly cleaned (including all holding containers) and good hygiene is undertaken to minimise the spread of disease and the risk of infestation of insects, rats and mice.
- The filtration system is maintained properly.
- Water purity is checked by chemical analysis using commercially available test kits.
- The facility has sterilisation measures to control bacteria/disease (e.g. UV lights, making sure bulbs are replaced regularly).
- Water temperature is measured regularly and always falls within the prescribed limits to maintain optimal health of the organisms.
- Adequate water flow is provided throughout the facility – a minimum of four times turnover an hour within the container for corals, with greater flow rates recommended (especially for corals such as Xenia).
- Animals are inspected at least daily.

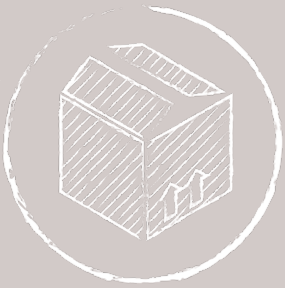
Corals should be placed so they do not touch each other – including making sure the corals' tentacles do not reach another coral at night.

Soft and hard corals should be kept in separate containers in the holding facility.

Temperature control

Maintain corals at the optimal temperature, which is below 80°F (26.7°C) for recirculating systems.

For flow through (open) systems, put intake pipe deep enough below the surface so that water temperature does not exceed 80°F (26.7°C).



Make sure all packing materials and shipping documents are accessible and filled out as much as possible before the pack starts.

Make sure that all packing materials and the packing area are clean.

Make sure hands are washed prior to packing, paying particular attention to remove/scrub away any oil or dirt and make sure to rinse hands well so no soap is left behind.

Do not pack corals before you have to, in order to ensure optimal health (i.e. do not pack animals earlier than needed).

Always screen organisms prior to export, making sure they are in good physical condition. Any animal that looks weak or is infected with parasites and bacteria should not be dispatched.

Keep physical contact with organisms to an absolute minimum.

Packing water should be uncontaminated – i.e. collected from a clean ocean site. Containers not in use should be covered at all times.

At a minimum, always double bag. For corals, more bags, thicker bags and clear liners are recommended.

Always only pack one live specimen/animal per bag.

For packing of aquarium organisms, carry out the following:

- Select a double bag and fill it with packing water to a level appropriate for the size of the organism to be placed into the bag;
- Gently place the organism into the bag.
- Make sure the animal is fully submerged.
- Take all the air out of the bag before adding oxygen.
- Add sufficient oxygen to the bag – make sure that the coral has sufficient water and oxygen.
- Twist the bag prior to sealing it – it is best practice to only twist the plastic that is to be banded/clipped and not the entire bag, but if you do then do so *slowly and gently*; if done quickly it may stress the animal inside or/and puncture the bag.
- Seal the bag tightly.
- Line a box with protective plastic liner to retain any potential leakage.
- Insulate the box (e.g. styrofoam box or if styrofoam is not available two boxes separated by crumpled, stuffed newspaper).

Corals should not be packed in water warmer than 80°F (26.7°C).

Always maintain bags upright.

Fill empty spaces in boxes with air-filled plastic bags.

If you are putting two layers of bags in a box, ensure that adequate support is provided for that second layer. In other words, ensure that the weight of the second layer is not resting on the first layer of bags, by placing a structure to hold up the second layer.

Ensure your shipment meets all the requirements prior to export, including but not limited to:

- 48-hour notification to importers;
- detailed packing list (quantities by species in each box);
- summary invoice;
- export permits; and
- copy of airway bill – with flight number and estimated time of arrival (ETA).

Label all boxes appropriately – species, number of animals, LIVE FISH, UP arrow, temperature, shipper's address and consignee address.

Always adhere to the International Air Transport Association's (IATA) and the specific airline's policies.

IATA specifies that corals should be packed for up to a 24-hour delay. If the departure flight is delayed by close to or more than 24 hours, all live animals will need to be brought back to the facility and repacked for the new flight schedule once it has been confirmed.

If the plane is delayed by less than 24 hours and the corals have not been packed to accommodate up to the 24-hour delay as recommended by IATA, the corals will need to be brought back to the facility and repacked for the new flight schedule.

Always keep boxes in a cool, shaded place. Do not store and/or transport packed boxes under the sun.

Total shipping time between exporter (beginning of pack) and importer (last specimen out of its bag) should ideally be 24 hours (up to 48 hours).

Always make sure to bring originals and copies of permits and other shipment documents to the airport.

If a connecting flight is needed between origin and destination, make sure that a backup flight to the same origin or destination is available within 24 hours of the original connecting flight – meaning your animals can get to your importer or sent back to you within 24 hours. If this is not the case, no live animals should be shipped to that destination.

ISBN 978-982-00-1193-9

Pacific Community

B.P. D5- 98848 Noumea Cedex
New Caledonia
Telephone: + 687 26 20 00
Email: spc@spc.int
Website: www.spc .int